

09/542121

=> d his

(FILE 'HOME' ENTERED AT 12:29:32 ON 13 MAR 2002)

INDEX 'ADISALERTS, ADISINSIGHT, ADISNEWS, AGRICOLA, ANABSTR, AQUASCI,  
BIOBUSINESS, BIOCOMMERCE, BIOSIS, BIOTECHABS, BIOTECHDS, BIOTECHNO,  
CABA,  
CANCERLIT, CAPLUS, CEABA-VTB, CEN, CIN, CONFSCI, CROPB, CROPUS, DDFB,  
DDFU, DGENE, DRUGB, DRUGLAUNCH, DRUGMONOG2, ...' ENTERED AT 12:29:46 ON  
13 MAR 2002

SEA (LACTIC(W)ACID(W)BACTERIA)

-----  
5 FILE ADISALERTS  
1 FILE ADISINSIGHT  
2\* FILE ADISNEWS  
2510 FILE AGRICOLA  
16 FILE ANABSTR  
106 FILE AQUASCI  
1112 FILE BIOBUSINESS  
76 FILE BIOCOMMERCE  
3974 FILE BIOSIS  
625 FILE BIOTECHABS  
625 FILE BIOTECHDS  
1352 FILE BIOTECHNO  
5952 FILE CABA  
62 FILE CANCERLIT  
6663 FILE CAPLUS  
446 FILE CEABA-VTB  
1 FILE CEN  
23 FILE CIN  
82 FILE CONFSCI  
1 FILE CROPB  
8 FILE CROPUS  
16 FILE DDFB  
9 FILE DDFU  
938 FILE DGENE  
16 FILE DRUGB  
2 FILE DRUGLAUNCH  
11 FILE DRUGU  
38 FILE EMBAL  
1203 FILE EMBASE  
1228 FILE ESBIOBASE  
35 FILE FOMAD  
24 FILE FOREGE  
5135 FILE FROSTI  
5585 FILE FSTA  
466 FILE GENBANK  
55 FILE HEALSAFE  
186 FILE IFIPAT  
1697 FILE JICST-EPLUS  
29 FILE KOSMET  
1994 FILE LIFESCI  
11 FILE MEDICONF  
1303 FILE MEDLINE  
1 FILE NIOSHTIC  
31 FILE NTIS  
27 FILE OCEAN  
4532 FILE PASCAL

1 FILE PHAR  
1 FILE PHIC  
23 FILE PHIN  
325 FILE PROMT  
4431 FILE SCISEARCH  
1222 FILE TOXCENTER  
547 FILE TOXLIT  
910 FILE USPATFULL  
1 FILE USPAT2  
1528 FILE WPIDS  
1528 FILE WPINDEX  
L1 QUE (LACTIC(W) ACID(W) BACTERIA)  
-----

FILE 'CAPLUS, CABA, FROSTI, FSTA, PASCAL, BIOSIS, SCISEARCH' ENTERED AT  
12:32:13 ON 13 MAR 2002

L2 441 S L1 AND (BETA-GALACTOSIDASE)  
L3 0 S L2 AND (4000(W) MILLER UNITS)  
L4 0 S L2 AND (MILLER UNITS)  
L5 0 S L2 AND MILLER  
L6 59670 S BETA-GALACTOSIDASE  
L7 53 S L6 AND (MILLER(W) UNITS)  
L8 0 S L7 AND (4000(W) MILLER)  
L9 19 DUP REM L7 (34 DUPLICATES REMOVED)

=> d 19 ibib ab 1-19

L9 ANSWER 1 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 1  
ACCESSION NUMBER: 2002:15400 CAPLUS  
TITLE: Measuring **.beta.-Galactosidase**  
Activity in Bacteria: Cell Growth, Permeabilization,  
and Enzyme Assays in 96-Well Arrays  
AUTHOR(S): Griffith, Kevin L.; Wolf, Richard E., Jr.  
CORPORATE SOURCE: Department of Biological Sciences, University of  
Maryland Baltimore County, Baltimore, MD, 21250, USA  
SOURCE: Biochemical and Biophysical Research Communications  
(2002), 290(1), 397-402  
CODEN: BBRCA9; ISSN: 0006-291X  
PUBLISHER: Academic Press  
DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB We describe a high-throughput procedure for measuring **.beta.-galactosidase** activity in bacteria. This procedure is unique because all manipulations, including bacterial growth and cell permeabilization, are performed in a 96-well format. Cells are permeabilized by chloroform/SDS treatment directly in the 96-well blocks and then transferred to 96-well microplates for std. colorimetric assay  
of **.beta.-galactosidase** activity as described by Miller [J. H. Miller (1972) Expts. in Mol. Genetics, Cold Spring Harbor Lab. Press, Cold Spring Harbor, NY]. Absorbance data are collected with a microplate reader and analyzed using a Microsoft Excel spreadsheet. The **.beta.-galactosidase** specific activity values obtained with the high-throughput procedure are identical to those obtained by the traditional single-tube method of Miller. Thus, values obtained with this procedure may be expressed as **Miller units** and compared directly to **Miller units** reported in the literature. The 96-well format for permeabilization and assay of enzyme specific activity together with the use of 12-channel and repeater pipettors enables efficient processing of hundreds of samples in an 8-h day. (c) 2002 Academic Press.  
REFERENCE COUNT: 7 THERE ARE 7 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE  
FORMAT

L9 ANSWER 2 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 2  
ACCESSION NUMBER: 2001:170031 CAPLUS  
DOCUMENT NUMBER: 135:267885  
TITLE: pAM401-based shuttle vectors that enable overexpression of promoterless genes and one-step purification of tag fusion proteins directly from *Enterococcus faecalis*  
AUTHOR(S): Fujimoto, Shuhei; Ike, Yasuyoshi  
CORPORATE SOURCE: Department of Microbiology School of Medicine, Gunma University, Gunma, 371-8511, Japan  
SOURCE: Appl. Environ. Microbiol. (2001), 67(3), 1262-1267  
CODEN: AEMIDF; ISSN: 0099-2240  
PUBLISHER: American Society for Microbiology  
DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB Two novel *Enterococcus faecalis*-*Escherichia coli* shuttle vectors that

utilize the promoter and ribosome binding site of bacA on the E. faecalis plasmid pPD1 were constructed. The vectors were named pMGS100 and pMGS101. PMGS100 was designed to overexpress cloned genes in E. coli and E. faecalis and encodes the bacA promoter followed by a cloning site and stop codon. PMGS101 was designed for the overexpression and purifn. of a cloned protein fused to a Strep-tag consisting of 9 amino acids at the carboxyl terminus. The Strep-tag provides the cloned protein with an affinity to immobilized streptavidin that facilitates protein purifn.

The

authors cloned a promoterless **.beta.-galactosidase** gene from E. coli and cloned the traA gene of the E. faecalis plasmid

pAD1

into the vectors to test gene expression and protein purifn., resp. **.beta.-Galactosidase** was expressed in E. coli and E. faecalis at levels of 103 and 10 **Miller units**, resp.

By cloning the pAD1 traA into pMGS101, the protein could be purified directly from a crude lysate of E. faecalis or E. coli with an immobilized

streptavidin matrix by one-step affinity chromatog. The ability of TraA to bind DNA was demonstrated by the DNA-assocd. protein tag affinity chromatog. method using lysates prep'd. from both E. coli and E. faecalis that overexpress TraA. The results demonstrated the usefulness of the vectors for the overexpression and cis/trans anal. of regulatory genes, purifn. and copurifn. of proteins from E. faecalis, DNA binding anal., detn. of translation initiation site, and other applications that require proteins purified from E. faecalis.

REFERENCE COUNT: 39 THERE ARE 39 CITED REFERENCES AVAILABLE FOR THIS

RECORD. ALL CITATIONS AVAILABLE IN THE RE

FORMAT

L9 ANSWER 3 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 3  
ACCESSION NUMBER: 2001:167568 CAPLUS  
DOCUMENT NUMBER: 134:339564  
TITLE: Characterization of an oxygen-dependent inducible promoter, the nar promoter of Escherichia coli, to utilize in metabolic engineering  
AUTHOR(S): Han, Se Jong; Chang, Ho Nam; Lee, Jongwon  
CORPORATE SOURCE: Department of Chemical Engineering and Bioprocess Engineering Research Center, Taejon, S. Korea  
SOURCE: Biotechnology and Bioengineering (2001), 72(5), 573-576  
CODEN: BIBIAU; ISSN: 0006-3592  
PUBLISHER: John Wiley & Sons, Inc.  
DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB The nar promoters, whose transcription is maximally induced under microaerobic conditions in the presence of nitrate ion, were characterized in fed-batch culture to det. whether they can be used for metabolic engineering, by which overall prodn. of valuable chems. can be increased. For this purpose, we tested whether the expression level of a reporter gene, the lacZ gene from the nar promoter, could be maintained const. throughout the induction period by manipulation of dissolved oxygen (DO) levels at a given nitrate ion concn. First, E. coli was grown under aerobic conditions (DO 80%) to absorbance at 600 nm (OD600) of 35, then the nar promoter was induced by redn. of DO to different levels, combined with different frequencies and duration of alternating microaerobic and aerobic conditions throughout the entire induction period. For a wild-type nar promoter (pMW61) in a mutant host E. coli with a mutation in the narG gene on the chromosome of the host (RK5265), it was possible to maintain prodn. of **.beta.-galactosidase** activity per cell (specific **.beta.-galactosidase** activity) at a const. rate at 5000, 10,000, 15,000, and 20,000 **Miller units**, using different combinations of nitrate ion concns. (0.1%,

0.5%, and 1%) and DO levels. In addn., it was possible to maintain prodn.

of specific  $\beta$ -galactosidase activity at a cons. rate at about 10,000 Miller units in the absence of nitrate ion when a nitrate-independent nar promoter (pMW618) in the narL-mutant of the W3110 E. coli strain (W3110narL-) was used. Based on these results, we conclude that the nar promoter system provides a convenient expression system for metabolic engineering as well as for maximal prodn. of recombinant proteins under conditions of fed-batch culture.

REFERENCE COUNT: 22 THERE ARE 22 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE REFORMAT

L9 ANSWER 4 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 4  
ACCESSION NUMBER: 2001:6607 CAPLUS  
DOCUMENT NUMBER: 134:126627  
TITLE: Coupling the T7 A1 promoter to the runaway-replication

vector as an efficient method for stringent control and high-level expression of lacZ

AUTHOR(S): Chao, Yun-Peng; Chern, Jong-Tzer; Wen, Chih-Sheng  
CORPORATE SOURCE: Department of Chemical Engineering, Feng Chia University, Taichung, Taiwan

SOURCE: Biotechnology Progress (2001), 17(1), 203-207  
CODEN: BIPRET; ISSN: 8756-7938

PUBLISHER: American Chemical Society

DOCUMENT TYPE: Journal

LANGUAGE: English

AB An expression vector characterized by tight regulation and high expression

of cloned genes appears to be indispensable for the engineering need. To achieve this goal, in assocn. with lacI the T7 A1 promoter contg. two synthetic lac operators was constructed into a runaway-replication vector.

To further examine this vector system, lacZ was subcloned and placed under

the control of the T7 A1 promoter on the plasmid. With the application of

the thermal induction alone, the Escherichia coli strain harboring the recombinant plasmid was able to produce 15,000 Miller units of  $\beta$ -galactosidase, while it yielded the recombinant protein with 45,000-50,000 Miller units upon both thermal and chem. induction. In sharp contrast, only 60-90 Miller units of  $\beta$ -galactosidase was obtained for the cell at an uninduced state. As a result, the prodn. yield of  $\beta$ -galactosidase over the background level is amplified approx. 170-fold by thermal induction and 500-fold by thermal and chem. induction. To produce the recombinant protein on a large scale, an approach by connecting two fermenters in series was newly developed. By applying the three-stage temp. shift in this dual fermenter system, 55,000 Miller units of  $\beta$ -galactosidase was obtained.

Overall, it shows the potential use of the vector system developed here for its tight control and high prodn. of recombinant proteins.

REFERENCE COUNT: 22 THERE ARE 22 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE REFORMAT

L9 ANSWER 5 OF 19 FSTA COPYRIGHT 2002 IFIS  
ACCESSION NUMBER: 2001(06):B0823 FSTA  
TITLE: Characterization of an oxygen-dependent inducible promoter, the nar promoter of Escherichia coli, to utilize in metabolic engineering.  
AUTHOR: Se Jong Han; Ho Nam Chang; Jongwon Lee

CORPORATE SOURCE:  
of

Correspondence (Reprint) address, Jongwon Lee, Dep.

Biochem., Sch. of Med., Catholic Univ. of  
Taegu-Hyosung, Taegu 705-034, Korea. Tel.  
+82-53-650-4471. Fax +82-53-621-4106. E-mail  
leejw(a)cuth.cataegu.ac.kr

SOURCE:

Biotechnology and Bioengineering, (2001) 72 (5)  
573-576, 22 ref.  
ISSN: 0006-3592

DOCUMENT TYPE:

Journal

LANGUAGE:

English

AB The O<sub>2</sub>-dependent nar promoter from *Escherichia coli*, which can be induced by lowering the dissolved O<sub>2</sub> (DO) concn. in the presence of nitrate ions, was studied to investigate its feasibility for use in metabolic engineering. Expression levels of the lacZ gene, encoding .beta.-galactosidase, under the control of the nar promoter were examined in *E. coli* grown initially under aerobic conditions

(80% DO) to an OD<sub>600</sub> of 35, followed by reduction of the DO

level. Using the wild-type nar promoter in *E. coli* containing a mutation in the narG gene (encoding a subunit of nitrate reductase), specific .beta.-galactosidase activity could be maintained at constant levels (5000-20 000 Miller units) using various nitrate ion concn. Using a nitrate-independent nar promoter in a narL<sup>+</sup> - *E. coli* mutant, specific .beta.-galactosidase activity could be maintained at a constant level of approx. 10 000 Miller units in the absence of nitrate ions. Results showed that the nar promoter system is a suitable expression system for metabolic engineering studies and for production of recombinant proteins in fed-batch culture.

L9 ANSWER 6 OF 19 CAPLUS COPYRIGHT 2002 ACS

DUPLICATE 5

ACCESSION NUMBER: 1998:301732 CAPLUS

DOCUMENT NUMBER: 129:77365

TITLE: Isolation and characterization of three *Streptococcus pneumoniae* transformation-specific loci by use of a lacZ reporter insertion vector

AUTHOR(S): Pestova, Ekaterina V.; Morrison, Donald A.

CORPORATE SOURCE: Laboratory for Molecular Biology, Department of Biological Sciences, University of Illinois at Chicago, Chicago, IL, USA

SOURCE: J. Bacteriol. (1998), 180(10), 2701-2710

CODEN: JOBAAY; ISSN: 0021-9193

PUBLISHER: American Society for Microbiology

DOCUMENT TYPE: Journal

LANGUAGE: English

AB Although more than a dozen new proteins are produced when *Streptococcus pneumoniae* cells become competent for genetic transformation, only a few of the corresponding genes have been identified to date. To find genes responsible for the prodn. of competence-specific proteins, a random lacZ transcriptional fusion library was constructed in *S. pneumoniae* by using the insertional lacZ reporter vector pEVP3. Screening the library for clones with competence-specific .beta.-galactosidase (.beta.-Gal) prodn. yielded three insertion mutants with induced .beta.-Gal levels of about 4, 10, and 40 Miller units.

In all three clones, activation of the lacZ reporter correlated with competence and depended on competence-stimulating peptide. Chromosomal loci adjacent to the integrated vector were subcloned from the insertion mutants, and their nucleotide sequences were detd. Genes at two of the loci exhibited strong similarity to parts of *Bacillus subtilis* com operons. One locus contained open reading frames (ORFs) homologous to

the comEA and comEC genes in *B. subtilis* but lacked a comEB homolog. A second locus contained four ORFs with homol. to the *B. subtilis* comG gene ORFs 1

to 4, but comG gene ORFs 5 to 7 were replaced in *S. pneumoniae* with an ORF encoding a protein homologous to transport ATP-binding proteins. Genes at all three loci were confirmed to be required for transformation by mutagenesis using pEVP3 for insertion duplications or an erm cassette for gene disruptions.

L9 ANSWER 7 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 6  
ACCESSION NUMBER: 1998:65317 CAPLUS  
DOCUMENT NUMBER: 128:214064  
TITLE: Modulation of the function of the signal receptor domain of XylR, a member of a family of prokaryotic enhancer-like positive regulators  
AUTHOR(S): Salto, Rafael; Delgado, Asuncion; Michan, Carmen; Marques, Silvia; Ramos, Juan L.  
CORPORATE SOURCE: Department of Biochemistry, Molecular and Cellular Biology of Plants, Estacion Experimental del Zaidin, Consejo Superior de Investigaciones Cientificas, Granada, E-18008, Spain  
SOURCE: J. Bacteriol. (1998), 180(3), 600-604  
CODEN: JOBAAY; ISSN: 0021-9193  
PUBLISHER: American Society for Microbiology  
DOCUMENT TYPE: Journal  
LANGUAGE: English

AB The XylR protein controls expression from the *Pseudomonas putida* TOL plasmid upper pathway operon promoter (Pu) in response to arom. effectors.

XylR-dependent stimulation of transcription from a Pu: lacZ fusion shows different induction kinetics with different effectors. With toluene, activation followed a hyperbolic curve with an apparent K of 0.95 mM and a

max.  $\beta$ -galactosidase activity of 2,550 Miller units. With o-nitrotoluene, in contrast, activation followed a sigmoidal curve with an apparent K of 0.55 mM and a Hill coeff. of 2.65. M-Nitrotoluene kept the XylR regulator in an inactive transcriptional form. Therefore, upon binding of an effector, the substituent on the arom. ring leads to productive or unproductive

XylR forms. The different transcriptional states of the XylR regulator are substantiated by XylR mutants. XylRE172K is a mutant regulator that is able to stimulate transcription from the Pu promoter in the presence of m-nitrotoluene; however, its response to m-aminotoluene was negligible, in contrast with the wild-type regulator. These results illustrate the importance of the electrostatic interactions in effector recognition and in the stabilization of productive and unproductive forms by the regulator

upon arom. binding. XylRD135N and XylRD135Q are mutant regulators that are able to stimulate transcription from Pu in the absence of effectors, whereas substitution of Glu for Asp135 in XylRD135E resulted in a mutant whose ability to recognize effectors was severely impaired. Therefore, the conformation of mutant XylRD135Q as well as XylRD135N seemed to mimic that of the wild-type regulator when effector binding occurred, whereas mutant XylRD135E seemed to be blocked in a conformation similar to that of

wild-type XylR and XylRE172K upon binding to an inhibitor mol. such as m-nitrotoluene or m-aminotoluene.

L9 ANSWER 8 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 7  
ACCESSION NUMBER: 1998:803860 CAPLUS  
DOCUMENT NUMBER: 130:205625  
TITLE: Development of a plasmid vector for overproduction of  $\beta$ -galactosidase in *Escherichia coli* by using genetic components of groEx from symbiotic bacteria in Amoeba proteus

AUTHOR(S) : Lee, Jung Eun; Ahn, Eun Young; Ahn, Tae In  
CORPORATE SOURCE: Department of Biology Education, Seoul National University, Seoul, 151-742, S. Korea  
SOURCE: J. Microbiol. Biotechnol. (1998), 8(5), 509-516  
CODEN: JOMBES; ISSN: 1017-7825  
PUBLISHER: Korean Society for Applied Microbiology  
DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB A plasmid vector, pXGPRMATTG-lac-Tgx, was developed for overprodn. of .  
**beta.-galactosidase** in Escherichia coli using the  
genetic components of groEx, a heat-shock gene cloned from symbiotic  
X-bacteria in Amoeba proteus. The vector is composed of intragenic  
promoters P3 and P4 of groEx, the structural gene of lac operon,  
transcription terminator signals of lac and groEx, and ColE1 and amp' of  
pBluescript SKII. The optimized host, E. coli DH5.alpha., transformed  
with the vector constitutively produced 117,310-171,961 Miller  
units of .**beta.-galactosidase** per mg protein  
in crude ext. The amt. of enzyme in crude ext. was 53% of total  
water-sol. proteins. About 43% of the enzyme could be purified to a  
specific activity of 322,249 Miller units/mg protein  
after two-fold purifn., using two cycles of pptn. with ammonium sulfate  
and one step of gel filtration. Thus, the expression system developed in  
this study presents a low cost and simple method for purifying  
overproduced .**beta.-galactosidase** in E. coli.

REFERENCE COUNT: 34 THERE ARE 34 CITED REFERENCES AVAILABLE FOR  
THIS

RECORD. ALL CITATIONS AVAILABLE IN THE RE  
FORMAT

L9 ANSWER 9 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 8  
ACCESSION NUMBER: 1998:486413 CAPLUS  
DOCUMENT NUMBER: 129:198611  
TITLE: Fed-batch cultivation of an oxygen-dependent  
inducible promoter system, the nar promoter in escherichia coli  
with an inactivated nar operon  
AUTHOR(S) : Han, Se Jong; Chang, Ho Nam; Lee, Jongwon  
CORPORATE SOURCE: Department of Chemical Engineering and Bioprocess  
Engineering Research Center, KAIST, Taejon, 305-701,  
S. Korea  
SOURCE: Biotechnol. Bioeng. (1998), 59(4), 400-406  
CODEN: BIBIAU; ISSN: 0006-3592  
PUBLISHER: John Wiley & Sons, Inc.  
DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB The nar promoter of Escherichia coli is maximally induced under anaerobic  
or microaerobic conditions in the presence of nitrate. We previously  
demonstrated in batch expts. that the intact nar promoter of E. coli  
cloned into a pBR322-based plasmid serves as a high-level expression  
system in a nar mutant of E. coli (J. Lee et al., 1996). In this study,  
we extend characterization of the nar promoter expression system to the  
fed-batch culture mode, which is widely used in industrial-scale fermn.  
From these expts., it was found that the specific .**beta.-galactosidase** activity expressed from the lacZ gene fused to the  
nar promoter was maximal when host cells were grown under aerobic  
conditions [dissolved oxygen, (DO) = 80%] to absorbance at 600 nm (OD600)  
= 35 before induction of the nar promoter by lowering DO to 1-2% with  
alternating microaerobic and aerobic conditions. Approx. 15 h after  
induction, the OD600 of the culture reached 135 and the specific .  
**beta.-galactosidase** activity increased to 40,000  
Miller units, equiv. to approx. 35% of the total  
cellular proteins. The specific 3-galactosidase activity before  
induction  
was approx. ,000 Miller units, giving an induction  
ratio of approx. 40. Based on these results, we conclude that the nar  
promoter provides a convenient and effective high level expression system

under conditions of fed-batch culture.

L9 ANSWER 10 OF 19 CAPLUS COPYRIGHT 2002 ACS  
ACCESSION NUMBER: 1996:661923 CAPLUS  
DOCUMENT NUMBER: 125:294710  
TITLE: Characterization of an oxygen-dependent inducible promoter system, the nar promoter, and Escherichia coli with an inactivated nar operon  
AUTHOR(S): Lee, Jintae; Cho, Moo Hwan; Lee, Jongwon  
CORPORATE SOURCE: Dep. Chem. Eng., Yeungnam Univ., Kyungbuk, S. Korea  
SOURCE: Biotechnol. Bioeng. (1996), 52(5), 572-578  
CODEN: BIBIAU; ISSN: 0006-3592

DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB The nar promoter of Escherichia coli, which is maximally induced under anaerobic conditions in the presence of nitrate, was characterized to see whether the nar promoter cloned onto pBR322 can be used as an inducible promoter. To increase the expression level, the nar promoter was expressed in E. coli where active nitrate reductase cannot be expressed from the nar operon on the chromosome. A plasmid with the lacZ gene expressing **.beta.-galactosidase** instead of the structural genes of the nar operon was used to simplify an assay of induction of the nar promoter. The following effects were investigated

to find optimal conditions: methods of inducing the nar promoter, optimal nitrate and molybdate concns. maximally inducing the nar promoter, the amt. of expressed **.beta.-galactosidase**, and induction ratio (specific **.beta.-galactosidase** activity after maximal induction/specific **.beta.-galactosidase** activity before induction.). The following results were obtained from

the expts.: induction of the nar promoter was optimal when E. coli was grown in the presence of 1% nitrate at the beginning of culture; expression of

**.beta.-galactosidase** was not affected by molybdate; the induction ratio was maximal, approx. 300, when the overnight culture was grown in the flask for 2.5 h (OD 1.3) before being transferred to the fermentor; the amt. of **.beta.-galactosidase** per cell and per medium vol. was maximal when E. coli was grown under aerobic conditions to OD = 1.7; then the nar promoter was induced under microaerobic conditions made by lowering dissolved oxygen level (DO) to 1-2%. After approx. 6 h of induction, OD600 became 3.2 and specific **.beta.-galactosidase** activity became 36,000

**Miller units**, equiv. to 35% of total cellular proteins, which was confirmed from SDS-PAGE.

L9 ANSWER 11 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 10  
ACCESSION NUMBER: 1996:112659 CAPLUS  
DOCUMENT NUMBER: 124:167135  
TITLE: Characterization of the nar promoter to use as an inducible promoter  
AUTHOR(S): Lee, Jintae; Cho, Moohwan; Hong, Eock-Kee; Kim, Kwang-Soo; Lee, Jongwon  
CORPORATE SOURCE: Dep. Chem. Eng., Yeungnam Univ., 705-034, S. Korea  
SOURCE: Biotechnol. Lett. (1996), 18(2), 129-34  
CODEN: BILED3; ISSN: 0141-5492

DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB The nar promoter of Escherichia coli was characterized, which is maximally induced under anaerobic conditions in the presence of nitrate. The following results are obtained; Expression of **.beta.-galactosidase** was optimal at 1% of nitrate and was not affected much by molybdate; the amt. of **.beta.-galactosidase** per unit vol. was maximal when the nar promoter was induced at OD600 = 1.7, and when anaerobic condition was made by supplying nitrogen gas. At

the optimal condition, the ratio of .beta.-galactosidase between before and after induction was approx. 250 **Miller units** were approx. 500. The results showed that the **tac** promoter can be used as an inducible promoter.

L9 ANSWER 12 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 11  
ACCESSION NUMBER: 1996:175149 CAPLUS  
DOCUMENT NUMBER: 124:222089  
TITLE: High-level expression of lacZ under control of the tac  
or trp promoter using runaway replication vectors in Escherichia coli  
AUTHOR(S): Kidwell, John; Kolibachuk, Dennis, Douglas  
CORPORATE SOURCE: Dep. Biol., James Madison Univ., Harrisonburg, VA, 22807, USA  
SOURCE: Biotechnol. Bioeng. (1996), 50(1), 108-14  
CODEN: BIBIAU; ISSN: 0006-3592  
DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB To det. the utility of coupling runaway replication to the expression of cloned genes under the control of strong promoters, lacZ transcriptional fusions to the trp or tac promoter (P<sub>trp</sub> or P<sub>tac</sub>) were constructed using plasmids in which the copy no. is thermally regulated. Cells contg. these plasmids were able to produce .beta.-galactosidase to levels between 3700 and 46,000 **Miller units** when induced only by a temp. upshift. The addn. of the appropriate chem. inducer, either IPTG (isopropyl-.beta.-D-thiogalactopyranoside) or IAA (3-.beta.-indoleacrylic acid), did not significantly enhance the thermal induction. The P<sub>tac</sub>-controlled and P<sub>trp</sub>-controlled lacZ induction differed slightly in that the P<sub>tac</sub>-controlled thermal induction exhibited a lag of approx. 1.5 h as compared to both chem. and thermal induction, whereas in the case of P<sub>trp</sub>-controlled induction, an increase in .beta.-galactosidase expression above background occurred at approx. the same time regardless of the means of induction. The best vector, a P<sub>trp</sub>-controlled lacZ fusion carried on a runaway replication vector having a basal copy no. of 10, was able to mediate the expression of .beta.-galactosidase to approx. 40,000 **Miller units** of .beta.-galactosidase comprising 25% of the total cell protein at 17 h postinduction under optimal conditions for protein yield. In these cells, lysis occurred as lacZ was maximally expressed. Under noninducing conditions, the plasmids were stable for at least 60 generations in the absence of antibiotic in batch culture.

L9 ANSWER 13 OF 19 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.  
ACCESSION NUMBER: 1997:87706 BIOSIS  
DOCUMENT NUMBER: PREV199799379419  
TITLE: The construction and use of promoter probe vectors for Rhodococcus sp.  
AUTHOR(S): Kayser, K. J.; Yun, C.-O.; Kilbane, J. J. II  
CORPORATE SOURCE: Inst. Gas Technol., 1700 S. Mt. Prospect Rd., Des Plaines, IL 60018 USA  
SOURCE: Actinomycetes, (1996) Vol. 7, No. 2, pp. 55-65.  
ISSN: 0732-0574.  
DOCUMENT TYPE: Article  
LANGUAGE: English  
AB Three promoter probe vectors have been constructed for use within Rhodococcus sp. They are hybrid replicons capable of replicating both in E. coli and Rhodococcus species due to the presence of replication functions derived from pUC19 and the Rhodococcus fascians plasmid pRF29 respectively. Promoter probe vector pRCM1 contains a promoterless gene which encodes a membrane-associated chloramphenicol efflux protein (cmr) derived from Rhodococcus fascians plasmid pRF2, pRCAT3 contains a promoterless chloramphenicol acetyl transferase gene (cat) derived from Tn9, and pEBC26 contains a promoterless **beta-**

galactosidase gene derived from pSVB-gal. Many derivatives of pRCM1 and pRCAT3, receiving inserts that regulated expression of chloramphenicol resistance in Rhodococcus sp. strain IGT8 proved to be unstable in E. coli, frequently yielding plasmids containing deletions. This instability was found to be largely associated with these vectors; however, some inserts of Rhodococcus DNA increased and others alleviated this instability. Derivatives of pEBC26 were stable both in Rhodococcus and E. coli and many DNA fragments encoding Rhodococcus promoters were isolated. The size of these promoter-containing DNA fragments ranged from 0.15 to 3 Kb and the level of beta-galactosidase expression in Rhodococcus hosts ranged from 0.1 to 838 Miller units. Promoters from Rhodococcus were not observed to function in E. coli; however, the E. coli rrnB promoter was shown to function weakly in Rhodococcus.

L9 ANSWER 14 OF 19 FSTA COPYRIGHT 2002 IFIS  
ACCESSION NUMBER: 1996(06):B0141 FSTA  
TITLE: High-level expression of lacZ under control of the tac  
or trp promoter using runaway replication vectors in Escherichia coli.  
AUTHOR: Kidwell, J.; Kolibachuk, D.; Dennis, D.  
CORPORATE SOURCE: Correspondence (Reprint) address, D. Dennis, Dep. of Biol., James Madison Univ., Harrisonburg, VA 22807, USA  
SOURCE: Biotechnology and Bioengineering, (1996) 50 (1) 108-114, 24 ref.  
ISSN: 0006-3592  
DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB The Escherichia coli lacZ gene, encoding .beta.-galactosidase, was placed under control of the trp or tac promoter in the runaway replication vectors pRA95 and pRA96, in which copy number is thermally regulated. Expression of lacZ was examined in transformed cells containing these plasmids. Increasing the temp. increased expression of the lacZ gene; 41.degree.C was the optimum temp. for thermal induction of gene expression. Induction of gene expression using isopropyl-.beta.-D-thiogalactopyranoside (IPTG) or 3-.beta.-indoleacrylic acid IAA did not significantly enhance thermal induction of gene expression. In thermally induced strains harbouring the tac promoter, a lag period of approx. 1.5 h was observed prior to .beta.-galactosidase production; no apparent lag was observed in strains possessing the trp promoter. Max. .beta.-galactosidase levels (up to 46 000 Miller units) were produced using a trp promoter on pRA96, having a basal copy number of 10; enzyme levels were 25% of the total cell protein 17 h after thermal induction.

L9 ANSWER 15 OF 19 FSTA COPYRIGHT 2002 IFIS  
ACCESSION NUMBER: 1997(02):B0111 FSTA  
TITLE: Characterization of an oxygen-dependent inducible promoter system, the nar promoter, and Escherichia coli with an inactivated nar operon.  
AUTHOR: Jintae Lee; Moo Hwan Cho; Jongwon Lee  
CORPORATE SOURCE: Correspondence (Reprint) address, Jongwon Lee, Dep. of Biochem., Sch. of Med., Catholic Univ. of Taegu-Hyosung, 3056-6, Daemyung 4-Dong, Nam-Gu, Taegu 705-034, Korea. Tel. 82-53-650-4471. Fax 82-53-621-4106  
SOURCE: Biotechnology and Bioengineering, (1996) 52 (5) 572-578, 22 ref.  
ISSN: 0006-3592  
DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB The nar promoter of Escherichia coli, which is optimally induced in the

presence of nitrate under anaerobic conditions, was characterized in order to ascertain its usefulness as an inducible promoter. The nar promoter was expressed in an E. coli strain having a mutant nar operon which does not express active nitrate reductase. A plasmid containing the lacZ gene, expressing .beta.-galactosidase, instead of the structural genes of the nar operon was used to assay induction of the nar promoter. Optimal conditions for nar induction were analysed. Results showed that induction of the nar promoter was optimal when E. coli was grown initially in the presence of 1% nitrate. Expression of the lacZ gene

was not affected by molybdate ions. The amount of .beta.-galactosidase per cell and per medium vol. was max. when E. coli was grown under aerobic conditions to an optical density (at 600 nm) of 1.7; induction of the nar promoter was observed by lowering dissolved O<sub>2</sub> concn. to microanaerobic levels (1-2%). After approx. 6 h induction, specific .beta.-galactosidase activity was 36 000 Miller units, equivalent to 35% of total cellular proteins, which was confirmed by SDS-PAGE. The specific activity of .beta.-galactosidase expressed from the nar promoter was comparable to that obtained using the tac and VHB promoters.

L9 ANSWER 16 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 12  
ACCESSION NUMBER: 1993:54760 CAPLUS  
DOCUMENT NUMBER: 118:54760  
TITLE: Experimental conditions may affect reproducibility of the .beta.-galactosidase assay  
AUTHOR(S): Giacomini, Alessio; Corich, Viviana; Ollero, Francisco  
CORPORATE SOURCE: J.; Squartini, Andrea; Nuti, Marco P.  
Padoua, Italy  
SOURCE: FEMS Microbiol. Lett. (1992), 100(1-3), 87-90  
CODEN: FMLED7; ISSN: 0378-1097  
DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB Several exptl. conditions and parameters contributing to the detn. of .beta.-galactosidase activity, as proposed in the J. H. Miller (1972) assay, were studied. This assay is based on bacterial cell permeabilization followed by spectrophotometric measurement of o-nitrophenol released from its galactoside conjugate. Use of the absorbance correction factor and the nature and concn. of permeabilizing agents were taken into account as different exptl. conditions. Reaction time, culture vol., and growth stage were investigated as equation parameters. From a quant. point of view the results, in terms of Miller units, are markedly affected by variation in these conditions. Therefore, to ensure reproducibility it is advisable to use const. values for all the parameters.

L9 ANSWER 17 OF 19 CAPLUS COPYRIGHT 2002 ACS  
ACCESSION NUMBER: 1990:546672 CAPLUS  
DOCUMENT NUMBER: 113:146672  
TITLE: Low copy number plasmids for regulated low-level expression of cloned genes in Escherichia coli with blue/white insert screening capability  
AUTHOR(S): Lerner, Claude G.; Inouye, Masayori  
CORPORATE SOURCE: Robert Wood Johnson Med. Sch., Rutgers, State Univ., Piscataway, NJ, 08854-5635, USA  
SOURCE: Nucleic Acids Res. (1990), 18(15), 4631  
CODEN: NARHAD; ISSN: 0305-1048  
DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB The authors constructed pCL1920 and pCL1921, a pair of low-copy-no. plasmids which contain a 580-bp BstUI fragment that carries the lac

promoter/operator, multiple cloning sites, and lacZ fragment of pUC19 cloned in place of the polylinker region in pGB2, a pSC101-derived plasmid

which confers spectinomycin (50  $\mu\text{g/mL}$ ) and streptomycin (100  $\mu\text{g/mL}$ ) resistance in *E. coli*. All multiple cloning sites indicated are unique except for an addnl. EcoRI site. Plasmids pCL19020 and pCL1921 contain the BstUI fragment in opposite orientations with respect to the pGB2 sequences. In the absence of inducer the pCL1920/21 vectors do not produce detectable levels of  $\beta$ -galactosidase in JM105 (lac<sup>q</sup> lacZ.DELTA.M15) cells (<2 Miller units). In the presence of 2 mM IPTG (isopropyl- $\beta$ -D-thiogalactopyranoside) the  $\beta$ -galactosidase levels of the pCL1920/21 [JM105] transformants rose to 11 units, whereas the pUC19 [JM105] transformants produced 470 units; a 43-fold increase. These results are consistent with the expected 40-fold difference in plasmid copy no. between pCL1920/21 (5 copies/cell) compared to that of the pUC vectors

(200

copies/cell). Thus, the pCL 1920 and pCL1921 vectors allow regulated low-level expression of genes inserted downstream of the lac promoter-operator when transformed into strains contg. the lac<sup>q</sup> gene. They should also be useful for cloning genes which may be deleterious at high copy no. Since the pCL1920/21 vectors are compatible with ColE-derived plasmids, they can be used to form stable co-transformants together with pBR322 or pUC-derived plasmids. For blue/white screening

of

inserts, competent host cells with the lacZ.DELTA.M15 gene are used, and the transformation mixt. is plated on LB, spectinomycin plates pre-spread with 5  $\mu\text{L}$  of 0.2 M IPTG and 25  $\mu\text{L}$  40 mg/mL 5-bromo-4-chloro-indolyl- $\beta$ -D-galactopyranoside per plate.

L9 ANSWER 18 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 13  
ACCESSION NUMBER: 1988:568937 CAPLUS  
DOCUMENT NUMBER: 109:168937  
TITLE: Fermentation of lactose by *Zymomonas mobilis* carrying a Lac+ recombinant plasmid  
AUTHOR(S): Yanase, Hideshi; Kurii, Junn; Tonomura, Kenzo  
CORPORATE SOURCE: Coll. Agric., Univ. Osaka Prefect., Osaka, 591, Japan  
SOURCE: J. Ferment. Technol. (1988), 66(4), 409-15  
CODEN: JFTED8; ISSN: 0385-6380  
DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB Lac+ recombinant plasmids encoding a  $\beta$ -galactosidase fused protein and lactose permease of *Escherichia coli* were introduced into *Z. mobilis*. The fused protein was expressed with 450 to 5860 Miller units of  $\beta$ -galactosidase activity, and functioned as lactase. Raffinose uptake by *Z. mobilis* CP4 was enhanced in the plasmid-carrying strain over the plasmid-free strain, suggesting that the lactose permease was functioning in the organism. *Z. mobilis* Carrying the plasmid could produce EtOH from lactose and whey, but could not grow on lactose as sole C source. Its growth was inhibited by either galactose or the galactose liberated from lactose.

L9 ANSWER 19 OF 19 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 14  
ACCESSION NUMBER: 1986:201248 CAPLUS  
DOCUMENT NUMBER: 104:201248  
TITLE: Construction of a new vector for the expression of foreign genes in *Zymomonas mobilis*  
AUTHOR(S): Byun, M. O. K.; Kaper, J. B.; Ingram, L. O.  
CORPORATE SOURCE: Dep. Microbiol. Cell Sci., Univ. Florida, Gainesville,  
FL, 32611, USA  
SOURCE: J. Ind. Microbiol. (1986), 1(1), 9-15  
CODEN: JIMIE7

DOCUMENT TYPE:

Journal

LANGUAGE:

English

AB Broad host range plasmids are suitable as vectors to introduce antibiotic resistance genes into *Z. mobilis*. However, attempts to use these vectors to carry other genes with enteric promoters and controlling elements have resulted in limited success due to poor expression. Thus, a promoter cloning vector was constructed in a modified pBR327 and used this vector to isolate 12 promoters from *Z. mobilis* which express various levels of ***beta*-galactosidase** in *Escherichia coli*. Four of these were then subcloned into pCVD 305 for introduction into *Z. mobilis*. All expressed ***beta*-galactosidase** in *Z. mobilis* with activities of 100-1800 Miller units. One of these retained a BamHI site into which new genes can be readily inserted immediately downstream from the *Z. mobilis* promoter. Genetic traits carried by pCVD 305 were initially unstable but spontaneous variants were produced during subculture in which the plasmid was resistant to curing

at

elevated temp. One of these variants was examd. The increased stability of this variant results from an alteration in the plasmid rather than a chromosomal mutation or from chromosomal integration.

=> d his

(FILE 'HOME' ENTERED AT 13:06:14 ON 13 MAR 2002)

INDEX 'ADISALERTS, ADISINSIGHT, ADISNEWS, AGRICOLA, ANABSTR, AQUASCI,  
BIOBUSINESS, BIOCOMMERCE, BIOSIS, BIOTECHABS, BIOTECHDS, BIOTECHNO,  
CABA,  
CANCERLIT, CAPLUS, CEABA-VTB, CEN, CIN, CONFSCI, CROPB, CROPU, DDFB,  
DDFU, DGENE, DRUGB, DRUGLAUNCH, DRUGMONOG2, ...' ENTERED AT 13:06:22 ON  
13 MAR 2002

SEA LACTOCOCCUS (W) LACTIS

-----  
3 FILE ADISALERTS  
1 FILE ADISINSIGHT  
3\* FILE ADISNEWS  
1417 FILE AGRICOLA  
7 FILE ANABSTR  
6 FILE AQUASCI  
526 FILE BIOBUSINESS  
12 FILE BIOCOMMERCE  
2620 FILE BIOSIS  
663 FILE BIOTECHABS  
663 FILE BIOTECHDS  
1535 FILE BIOTECHNO  
4137 FILE CABA  
40 FILE CANCERLIT  
2854 FILE CAPLUS  
321 FILE CEABA-VTB  
1 FILE CEN  
9 FILE CIN  
41 FILE CONFSCI  
12 FILE DDFU  
1111 FILE DGENE  
2 FILE DRUGNL  
15 FILE DRUGU  
2 FILE DRUGUPDATES  
16 FILE EMBAL  
1463 FILE EMBASE  
1157 FILE ESBIOBASE  
1 FILE FOREGE  
1004 FILE FROSTI  
1691 FILE FSTA  
1211 FILE GENBANK  
4 FILE HEALSAFE  
72 FILE IFIPAT  
262 FILE JICST-EPLUS  
19 FILE KOSMET  
1563 FILE LIFESCI  
1839 FILE MEDLINE  
1 FILE NIOSHTIC  
3 FILE NTIS  
1746 FILE PASCAL  
1 FILE PHAR  
4 FILE PHIN  
39 FILE PROMT  
2800 FILE SCISEARCH  
630 FILE TOXCENTER  
768 FILE TOXLIT

299 FILE USPATFULL  
1 FILE SPAT2  
159 FILE PIDS  
159 FILE WPINDEX  
L1 QUE LACTOCOCCUS (W) LACTIS  
-----

FILE 'CABA, CAPLUS, SCISEARCH, BIOSIS' ENTERED AT 13:07:27 ON 13 MAR 2002  
L2 84 S L1 AND PERMEAB?  
L3 45 DUP REM L2 (39 DUPLICATES REMOVED)

L3 ANSWER 36 OF 45 CAPLUS COPYRIGHT 2002 ACS DUPLICATE 17  
ACCESSION NUMBER: 1992:586759 CAPLUS  
DOCUMENT NUMBER: 117:186759  
TITLE: Effect of the unsaturation of phospholipid acyl chains  
on leucine transport of *Lactococcus lactis* and membrane permeability  
AUTHOR(S): In 't Veld, Gerda; Driessen, Arnold J. M.; Konings, Wil N.  
CORPORATE SOURCE: Dep. Microbiol., Univ. Groningen, Haren, 9751 NN, Neth.  
SOURCE: Biochim. Biophys. Acta (1992), 1108(1), 31-9  
CODEN: BBACAO; ISSN: 0006-3002  
DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB The effect of the degree of unsatn. of the phospholipid acyl chains on the branched-chain amino acid transport system of *L. lactis* was investigated by the use of a membrane fusion technique. Transport activity was analyzed in hybrid membranes composed of equimolar mixts. of synthetic unsatd. phosphatidylenthanolamine (PE) and phosphatidylcholine (PC) in which the no. of cis double bonds in the 18-carbon acyl chains was varied.

The accumulation level and initial rate of both counterflow and protonmotive-force driven transport of leucine decreased with increasing no. of double bonds. The redn. in transport activity with increasing no. of double bonds correlated with an increase in the passive permeability of the membranes to leucine. The membrane fluidity was hardly affected by the double bond content. It is concluded that the degree of lipid acyl chain unsatn. is a minor determinant of the activity of the branched chain amino acid transport system, but effects strongly the passive permeability of the membrane.

L3 ANSWER 37 OF 45 CABA COPYRIGHT 2002 CABBI DUPLICATE 18  
ACCESSION NUMBER: 92:1627 CABA  
DOCUMENT NUMBER: 920449928  
TITLE: The bacteriocin lactococcin A specifically increases permeability of lactococcal cytoplasmic membranes in a voltage-independent, protein-mediated manner  
AUTHOR: Belkum, M. J. van; Kok, J.; Venema, G.; Holo, H.; Nes, I. F.; Konings, W. N.; Abeel, T.; Van Belkum, M.  
CORPORATE SOURCE: Department of Genetics, University of Groningen, Kerklaan 30, 9751NN Haren, Netherlands.  
SOURCE: Journal of Bacteriology, (1991) Vol. 173, No. 24, pp. 7934-7941. 36 ref.  
ISSN: 0021-9193  
DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB Lactococcin A is a bacteriocin produced by *Lactococcus lactis*. Its structural gene has recently been cloned and sequenced. Purified lactococcin A increased the permeability of the cytoplasmic membrane of *L. lactis* and dissipated the membrane potential. A significantly higher concentration of lactococcin A was needed to dissipate the membrane potential in an immune strain of *L.*

*lactis*. Lactococcin A at low concentrations (0.029 micro g/mg of protein) inhibited secondary and phosphate-bond driven transport of amino acids in sensitive cells and caused efflux of preaccumulated amino acids.

Accumulation of amino acids by immune cells was not affected by this concentration of lactococcin A. Lactococcin A also inhibited proton motive force-driven leucine uptake and leucine counterflow in membrane vesicles of the sensitive strain but not in membrane vesicles of the immune strain.

These observations indicate that lactococcin A makes the membrane **permeable** for leucine in the presence or absence of a proton motive force and that the immunity factor(s) is membrane linked. Membrane vesicles of *Clostridium acetobutylicum*, *Bacillus subtilis* and *Escherichia coli* were not affected by lactococcin A, nor were liposomes derived from phospholipids of *L. lactis*. These results indicate that lactococcin A acts

on the cytoplasmic membrane and is very specific towards lactococci. The combined results obtained with cells, vesicles and liposomes suggest that the specificity of lactococcin A may be mediated by a receptor protein associated with cytoplasmic membrane.

L3 ANSWER 38 OF 45 SCISEARCH COPYRIGHT 2002 ISI (R)

ACCESSION NUMBER: 91:688106 SCISEARCH

THE GENUINE ARTICLE: GV187

TITLE: THE BACTERIOCIN LACTOCOCCIN-A SPECIFICALLY INCREASES  
PERMEABILITY OF LACTOCOCCAL CYTOPLASMIC MEMBRANES  
IN A VOLTAGE-INDEPENDENT, PROTEIN-MEDIATED MANNER

AUTHOR: VANBELKUM M J (Reprint); KOK J; VENEMA G; HOLO H; NES I  
F;

CORPORATE SOURCE: KONINGS W N; ABEE T

UNIV GRONINGEN, DEPT GENET, KERKLAAN 80, 9751 NN HAREN,  
NETHERLANDS (Reprint); NLVF, MICROBIAL GENE TECHNOL LAB,  
N-1432 AS, NORWAY; UNIV GRONINGEN, DEPT MICROBIOL, 9751

NN

COUNTRY OF AUTHOR: HAREN, NETHERLANDS

NETHERLANDS; NORWAY

SOURCE: JOURNAL OF BACTERIOLOGY, (1991) Vol. 173, No. 24, pp.  
7934-7941.

DOCUMENT TYPE: Article; Journal

FILE SEGMENT: LIFE

LANGUAGE: ENGLISH

REFERENCE COUNT: 36

\*ABSTRACT IS AVAILABLE IN THE ALL AND IALL FORMATS\*

AB Lactococcin A is a bacteriocin produced by *Lactococcus* *lactis*. Its structural gene has recently been cloned and sequenced (M. J. van Belkum, B. J. Hayema, R. E. Jeeninga, J. Kok, and G. Venema, Appl. Environ. Microbiol. 57:492-498, 1991). Purified lactococcin A increased the **permeability** of the cytoplasmic membrane of *L. lactis* and dissipated the membrane potential. A significantly higher concentration of lactococcin A was needed to dissipate the membrane potential in an immune strain of *L. lactis*. Lactococcin A at low concentrations (0.029-mu-g/mg of protein) inhibited secondary and phosphate-bond driven transport of amino acids in sensitive cells and caused efflux of preaccumulated amino acids. Accumulation of amino acids by immune cells was not affected by this concentration of lactococcin A. Lactococcin A also inhibited proton motive force-driven leucine uptake

and

leucine counterflow in membrane vesicles of the sensitive strain but not in membrane vesicles of the immune strain. These observations indicate that lactococcin A makes the membrane **permeable** for leucine in the presence or absence of a proton motive force and that the immunity factor(s) is membrane linked. Membrane vesicles of *Clostridium acetobutylicum*, *Bacillus subtilis*, and *Escherichia coli* were not affected by lactococcin A, nor were liposomes derived from phospholipids of *L. lactis*. These results indicate that lactococcin A acts on the cytoplasmic membrane and is very specific towards lactococci. The combined results

obtained with cells, vesicles, and liposomes suggest that the specificity of lactococcin A be mediated by a receptor protein associated with the cytoplasmic membrane.

L3 ANSWER 39 OF 45 CABA COPYRIGHT 2002 CABI                   DUPLICATE 19  
ACCESSION NUMBER:   91:11466 CABA  
DOCUMENT NUMBER:   910443783  
TITLE:   Characterization of lactococci and lactobacilli isolated from semihard goats' cheese  
AUTHOR:    Requena, T.; Pelaez, C.; Desmazeaud, M. J.  
CORPORATE SOURCE:    Instituto del Frio (CSIC), Ciudad Universitaria, 28040 Madrid, Spain.  
SOURCE:   Journal of Dairy Research, (1991) Vol. 58, No. 1, pp. 137-145. 32 ref.  
ISSN: 0022-0299  
DOCUMENT TYPE:   Journal  
LANGUAGE:    English  
AB Several strains of *Lactococcus lactis* var. *lactis*, *Lactobacillus casei* and *Lactobacillus plantarum* isolated from traditional goat milk cheese were studied for titratable acidity, proteolysis in milk and enzymic activities. Aminopeptidase activities were measured with whole cells and cells made **permeable** with Triton X-100. Caseinolytic activity was investigated using PAGE. *Lc. lactis* var. *lactis* had a level of proteolytic activity in skim milk greater than that of *Lb. casei*, while this activity in *Lb. plantarum* was very low. Alanine aminopeptidase activity was almost non-existent for all strains tested, while lysine aminopeptidase activity appeared to be of fundamentally intracellular origin. Leucine aminopeptidase activity was also greater in **permeable** cells than in whole cells for *Lb. casei* and *Lb. plantarum*. *Lc. lactis* var. *lactis* leucine aminopeptidase activity was greater in whole cells. No significant hydrolysis of casein was found with *Lb. casei* IFPL 725 and *Lb. plantarum* IFPL 722 made **permeable** with Triton X-100 after 24 h incubation with whole bovine casein.

L3 ANSWER 40 OF 45 CABA COPYRIGHT 2002 CABI  
ACCESSION NUMBER:   89:65434 CABA  
DOCUMENT NUMBER:   890432541  
TITLE:   Characterization of an aminopeptidase of *Streptococcus cremoris* AM2 and an alpha -galactosidase of *Leuconostoc lactis* CNRZ 1091  
AUTHOR:    Boquien, C. Y.; Desmazeaud, M. J.; Corrieu, G.  
CORPORATE SOURCE:    INRA, Lab. Genie des Procedes Biotech. Agro-alimentaires, 78850 Thiverval-Grignon, France.  
SOURCE:   Lait, (1989) Vol. 69, No. 1, pp. 71-81. 21 ref.  
DOCUMENT TYPE:   Journal  
LANGUAGE:    French  
SUMMARY LANGUAGE:    English  
AB The API ZYM enzyme system was used to detect enzymic activities specific to lactic acid bacteria. A total of 20 mesophilic streptococcal and 34 leuconostoc strains were tested, and 2 enzymes selected for characterization were (i) an aminopeptidase of *Streptococcus cremoris* AM2 that hydrolysed histidyl-phenylalanine- beta -naphthylamide and (ii) an alpha -galactosidase of *Leuconostoc lactis* CNRZ 1091 that hydrolysed paranitrophenol- alpha -galactose. Activity of (i) was max. in fresh cells whilst that of (ii) was max. when the cells were **permeabilized** with Triton X-100. Enzymes (i) and (ii) resp. had pH optimum of 7 and 6.5, and optimum temp. was 40 deg C for both enzymes. Apparent Michaelis

constant (Km) was 0.17 and 0.73 mM for (i) and (ii) resp., and Vmax was 0.6 and 90 pmol/s or 107 c.f.u. per ml.

L3 ANSWER 41 OF 45 CABA COPYRIGHT 2002 CABI  
ACCESSION NUMBER: 84:85357 CABA  
DOCUMENT NUMBER: 840491300  
TITLE: Fermentation process improvement by membrane technology  
AUTHOR: Kyung, K. H.  
CORPORATE SOURCE: Michigan State Univ., East Lansing, Michigan 48824,  
USA.  
SOURCE: Dissertation Abstracts International, B (Sciences and Engineering), (1984) Vol. 44, No. 7, pp. 2071-2072.  
Order No: DA8324736.  
DOCUMENT TYPE: Journal  
LANGUAGE: English

AB For continuous fermentation of glucose to ethanol using *Saccharomyces cerevisiae* ATCC 4126, substrate was fed into a continuous dialysate circuit, then transferred by membrane diffusion through an intermediate dialyser into a batch fermentor circuit; product was simultaneously withdrawn from the fermentor circuit through dialyser membranes into the dialysate circuit and out in the effluent. Cells in the fermentor were essentially immobilized, and converted substrate to product by maintenance metabolism. Advantages of the system were offset by low productivity which was apparently limited by membrane **permeability**. A batch fermentation system utilizing a mutualistic dialysis culture of *Streptococcus lactis* and *Candida utilis* was also investigated, the 2 organisms being inoculated into separate fermentors connected by an intermediate dialyser. Lactose was fermented by *S. lactis* to lactic acid which was dialysed into the *C. utilis* culture and utilized to produce yeast biomass. Solute exchange rate across the membrane was the primary limiting factor.

L3 ANSWER 42 OF 45 CABA COPYRIGHT 2002 CABI  
ACCESSION NUMBER: 84:76732 CABA  
DOCUMENT NUMBER: 840491084  
TITLE: Location of peptidases outside and inside the membrane of *Streptococcus cremoris*  
AUTHOR: Exterkate, F. A.  
CORPORATE SOURCE: Netherlands Inst. for Dairy Res., NIZO, Ede,  
Netherlands.  
SOURCE: Applied and Environmental Microbiology, (1984) Vol. 47, No. 1, pp. 177-183. 24 ref.  
ISSN: 0099-2240  
DOCUMENT TYPE: Journal  
LANGUAGE: English

AB Peptidase activity determinations involving native cells of *Streptococcus cremoris* and completely disrupted cell preparations, as well as experiments concerned with peptidase activity distribution among cell fractions obtained by a damage-restrictive removal of the cell wall and release of intracellular material, suggest the presence of peptidases with distinguishable locations. Alanyl, leucyl and prolyl aminopeptidase activities are most likely located in the cell wall-membrane interface, showing no detectable association with the membrane. Lysyl aminopeptidase is present not only in this location, but also as an intracellular enzyme. Endopeptidase activity and glutamate aminopeptidase activity appear to be weakly associated with the membrane. Results of experiments concerned with permeabilization of the membrane, and findings regarding an effect of the local environment of the enzymes on their pH activity profiles are evaluated and considered as being indicative of the proposed location.

The

possible implications of these findings with respect to protein utilization during growth of the organism in milk discussed.

L3 ANSWER 43 OF 45 CABA COPYRIGHT 2002 CABI  
ACCESSION NUMBER: 82:16001 CABA  
DOCUMENT NUMBER: 810469483  
TITLE: Regulation of methyl- beta  
-D-thiogalactopyranoside-  
AUTHOR: 6-phosphate accumulation in Streptococcus lactis by exclusion and expulsion mechanisms  
CORPORATE SOURCE: Thompson, J.; Saier, M. H., Jr.  
Dep. of Biol., John Muir Coll., Univ. of California at San Diego, La Jolla, California 92093, USA.  
SOURCE: Journal of Bacteriology, (1981) Vol. 146, No. 3,  
PP. 885-894. 40 ref.  
ISSN: 0021-9193  
DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB Starved cells of Streptococcus lactis ML3 (grown previously on galactose, lactose or maltose) accumulated methyl- beta -D-thiogalactopyranoside (TMG) by the lactose:phosphotransferase system. >98% of accumulated sugar was present as a phosphorylated derivative, TMG-6-phosphate (TMG-6P).

When a phosphotransferase system sugar (glucose, mannose, 2-deoxyglucose, or lactose) was added to the medium simultaneously with TMG, the beta -galactoside was excluded from the cells. Galactose enhanced the accumulation of TMG-6P. Glucose, mannose, lactose, or maltose + arginine, when added to a suspension of TMG-6P-loaded cells of S. lactis ML3, elicited rapid expulsion of intracellular solute. The material recovered in the medium was exclusively free TMG. Expulsion of galactoside required both entry and metabolism of an appropriate sugar, and intracellular dephosphorylation of TMG-6P preceded efflux of TMG. The rate of dephosphorylation of TMG-6P by permeabilized cells was increased 2-3X by adenosine 5'-triphosphate but was strongly inhibited by fluoride. S. lactis ML3 (DGr) was derived from S. lactis ML3 by positive selection for resistance to 2-deoxy-D-glucose and was defective in the enzyme IIMan component of the glucose:phosphotransferase system. Neither glucose nor mannose excluded TMG from cells of S. lactis ML3 (DGr), and these 2 sugars

failed to elicit TMG expulsion from preloaded cells of the mutant strain. Accumulation of TMG-6P by S. lactis ML3 can be regulated by two independent mechanisms whose activities promote exclusion or expulsion of galactoside from the cell.

L3 ANSWER 44 OF 45 CABA COPYRIGHT 2002 CABI  
ACCESSION NUMBER: 82:19154 CABA  
DOCUMENT NUMBER: 820474332  
TITLE: Membrane-bound peptidases in Streptococcus cremoris  
AUTHOR: Exterkate, F. A.; International Dairy Federation,  
Symposium  
CORPORATE SOURCE: Netherlands Inst. for Dairy Res. (NIZO), Ede,  
Netherlands.  
SOURCE: Netherlands Milk and Dairy Journal, (1981) Vol. 35,  
No. 3/4, pp. 328-332. 12 ref.  
ISSN: 0028-209X  
DOCUMENT TYPE: Journal  
LANGUAGE: English  
AB Review of results of various experiments, relating to peptidase activity distribution among cell fractions, permeabilization of the membrane, and initial reaction kinetics, leads to the conclusion that Streptococcus cremoris has a functional system of specific peptidases located within the membrane and at its outside surface. This system plays a role in protein utilization during growth of the organism in milk, by making small peptides and free amino acids available for transport through

the membrane.

L3 ANSWER 45 OF 45 CABA COPYRIGHT 2002 CABI  
ACCESSION NUMBER: 80:20598 CABA  
DOCUMENT NUMBER: 800458595  
TITLE: Effect of external factors on manifestation of  
antagonistic activity by lactic streptococci  
AUTHOR: Zaborskikh, E. I.  
SOURCE: Biologiya mikroorganizmov i ikh ispol'zovanie v  
narodnom khozyaistve, (1977) pp. 99-103, 208. 10  
ref.  
Publisher: Gosudarstvennyi Universitet imeni A. A.  
Zhdanova. Irkutsk  
PUB. COUNTRY: USSR  
DOCUMENT TYPE: Miscellaneous  
LANGUAGE: Russian  
AB Cultures of 2 strains of *Streptococcus lactis* and 2 of *Str. diacetilactis*  
showing antagonism against *Escherichia coli* were filtered through  
membrane  
filters; the filtrates and hydrolysed milk as control were inoculated  
with  
E. coli at 20 to 2 X 10<sup>8</sup> cells/ml, and growth intensity measured by  
nephelometer after incubation for 24 h at 30 deg C. The filtrates  
inhibited E. coli growth irrespective of size of inoculum. The  
antimicrobial compounds formed by the strains studied were stable to  
heating for 30 min at 65 deg C or to boiling for 20 min; they retained  
their activities in acid, but were inactivated under alkaline conditions;  
they passed through semi-permeable membranes; they were not  
precipitated by trichloroacetic acid, saturated (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub> solution,  
ammonia or ethanol; and they were not extractable by diethyl ether.